

ELECTROPHYSIOLOGICAL MEASUREMENTS OF CARBON
DIOXIDE PERCEPTION IN THREE SPECIES
OF TICKS (ACARINA: IXODIDAE)

By

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DIOXIDE PERCEPTION IN THREE SPECIES
OF TICKS (ACARINA: IXODIDAE)

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PREFACE

This investigation was designed to determine if certain physiological or environmental factors influence carbon dioxide perception in Amblyomma maculatum Koch, Amblyomma americanum (L.) and Dermacentor variabilis (Say). Actual measurement of carbon dioxide perception was accomplished through use of electrophysiological techniques. The effects of species, sex, age, humidity, temperature and ambient CO₂ concentration on carbon dioxide perception were determined and statistically analyzed.

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CHAPTER I

INTRODUCTION

Tick chemoattraction has received substantial attention among researchers in recent years and much emphasis has been directed towards its possible utilization in tick control programs and population density studies. To date, the majority of work in this area has dealt with using carbon dioxide (CO₂) as the attracting medium. Garcia (1962, 1965, 1969), Nevill (1964) and Miles (1968) utilized CO₂ to trap significant numbers of several different tick species. Wilson et al. (1972) incorporated CO₂ in the form of dry ice to collect all developmental stages of the lone star tick, Amblyomma americanum (L.). A small percentage of marked adults were drawn to the CO₂ source from a maximum distance of 21.3 m.

Laboratory studies by Sauer et al. (1974) on A. americanum further indicated that the intensity of tick responses to CO₂ is dependent on CO₂ concentration. Although these investigations give support to CO₂ being a stimulant or attractant of ticks, results obtained have been inconsistent (Gearhart et al., unpublished).

Indications are that other physiological or environmental factors may play a significant role in tick perception of CO₂. The objective of this study was to determine if differences in species, sex, age, humidity, temperature and ambient CO₂ concentrations had an effect on electrical activity of nerve cells in the Haller's organ in three species of

ixodid ticks.

Evaluation of these parameters was accomplished through electrophysiological techniques. Research results of this type, as they apply to ticks, are sparse. However, work of a somewhat similar nature has been conducted on various species of Diptera (Agee, 1973 and Davis, 1976a, 1976b), Lepidoptera (Baker and Roelofs, 1976), Isoptera (Floyd et al., 1976) and Orthoptera (Washio and Nishino, 1976) and will serve as a reference for experimental designs and techniques in these studies.

CHAPTER II

REVIEW OF LITERATURE

The fact that ticks are capable of sensing and reacting to various chemical stimuli has been researched and documented for many years. LaHille (1905) was the first investigator to postulate that the receptors responsible for olfactory perception are located in Haller's organ. This structure, located on the distal portion of tarsus I of the foreleg, had previously been thought to be auditory in function. In his work LaHille found that normal Boophilus annulatus would not walk across a strip of paper impregnated with Sanol. However, if these particular ticks had their Haller's organs amputated they would not hesitate in their movement across this barrier. Behavioral studies on the sensory perceptions of Argas persicus (Oken) (Hindle and Merriman, 1912), Ixodes ricinus (L.) (Lees, 1948) and Amblyomma hebraeum Koch (Rechav et al., 1977) and electrophysiological studies on the sensory physiology of Ixodes persulcatus P. Sch. (Zolotarev and Elizarov, 1963) have since added support to this original theory of Haller's organ involvement in olfactory perception.

Morphologically, Nuttall (1908) first determined that the Haller's organs of ixodid ticks are comprised of two distinct parts, which he referred to as the anterior pit and posterior capsule. However, it was not until the advent of techniques such as scanning electron microscopy (SEM) that a true understanding of the structure and morphology of this

organ became known. Bruce (1971) utilized SEM to study both the external and internal features of the posterior capsule of Haller's organ in the lone star tick, Amblyomma americanum (L.). Externally be observed a well defined horizontal slit or aperture located in the thin membranous cuticle covering the capsule, with variation occurring between right and left legs as well as between sexes. This concept of external variation in Haller's organ was also observed by Homsher and Sonenshine (1975) in their studies of ten species of ixodid ticks and by Chow and Wang (1975) in investigations of the ixodid ticks Rhipicephalus sanguineus (Latreilla), Amblyomma testudinarium Koch and Boophilus microplus (Canastrini). Internally, Bruce observed several irregularly shaped pleomorphs and cylindrical spines, along with eight symmetrically arranged thin-walled setae. No internal variations were observed between either right or left legs or between sexes.

A more detailed account of the ultrastructure of Haller's organ in A. americanum was conducted by Foelix and Axtell (1972). Their studies, which utilized SEM, included observations of both the anterior pit and posterior capsule and agreed with the findings of Bruce with one exception. Whereas Bruce found eight thin-walled setae in the posterior capsule, Foelix and Axtell could only demonstrate seven.

Scanning electron microscopy studies have also been carried out by Chu-Wang and Axtell (1973) on the claw sensilla of the soft tick Argas arboreus Kaiser, Hoogstraal and Kohls and the hard tick A. americanum and by Waladde (1977) on the cattle tick, B. microplus. They found these sensilla to function not only as mechanoreceptors but as chemoreceptors also.

In her review of the structure of arthropod chemoreceptors, Slifer

(1970) notes that a typical chemoreceptor is composed of (a) cuticular parts, (b) sensory neurons and (c) sheath cells. The cuticular parts are subdivided into thick-walled and thin-walled chemoreceptors, such as those observed by Bruce in the Haller's organ of A. americanum.

Several authors have studied the neural mechanisms which control arthropod behavior and olfactory perception (Erickson, 1963; Huber, 1967; Roeder, 1970; Kaissling, 1971 and Balashov, 1972). In Balashov's work on several species of Ixodidae, he found that ticks have retained a simple bineural (reflex) arc consisting of sensory and motor or association motor neurons from several pairs of glomerular nuclei, which lie within the ganglia of leg I and are connected to Haller's organ. These allow the tick to perceive odors from a considerable distance. Balashov found this particular arrangement to be specific for ixodid ticks.

Behavioral studies by Lees (1948) on the sensory physiology of the sheep tick, I. ricinus, indicate that attraction to a host is due primarily to odors being emitted by the host and temperature stimulation. In regards to temperature, Lees was able to show that ticks are attracted or repelled by changes in air temperature and not by radiant heat. Similar investigations performed by Wilkinson (1953) on the larvae of the cattle tick, B. microplus, agreed with the findings of Lees. More specifically, on responses to odor, Wilkinson demonstrated that ticks were greatly attracted to a test tube rubbed in human sweat and to cotton swabs which had been rubbed in mucus, saliva and areas of the flank and escutcheon of cattle.

Tick attraction resulting from odors being emitted by the host has also been substantiated by Dethier (1957) in his review of the blood

sucking arthropods and Balashov (1972) in his work with several species of ixodid ticks. Balashov showed that ticks which were released at varying distances from a frequently used host path would orient themselves in close proximity to that path for periods as long as one month after release. He concluded, therefore, that attraction to the path was due to host odor.

Concerning physiological factors which may influence host-seeking behavior, Sactor et al. (1948) calculated the optimum temperature for molting in laboratory reared A. americanum to be 30°C. Relative humidity of 85-100% was also proven to be most favorable for attachment and engorgement. This finding coincides with that of Sauer and Hair (1971) who showed the critical equilibrium humidity (CEH) for A. americanum to be 85% for females and slightly less for males. Hair et al. (1975) made comparisons between the abilities of A. americanum, Amblyomma maculatum Koch and Dermacentor variabilis (Say) to resist dehydration and maintain water balance. Of the three species, A. americanum appeared most dependent on moist microenvironments because of its greater susceptibility to loss of total body water and drop of hemolymph at low humidities.

Although investigations, such as those previously mentioned, have indicated that ticks are attracted to certain volatile chemical odors being emitted by the host, limited attempts have been taken towards the discovery, identification and utilization of these particular compounds. Only carbon dioxide (CO₂) has been investigated to any great degree as a possible tick stimulant for host-seeking behavior.

Garcia (1962, 1965, 1969) utilized CO₂ to trap significant numbers of Ornithodoros coriaceus Koch, Dermacentor occidentalis Marx, Ixodes

pacificus Cooley and Kohls and Dermacentor andersoni Stiles in both laboratory and field tests. Two other members of the genus Ornithodoros have also been collected with the aid of CO₂ (Nevill, 1964 and Miles, 1968). Nevill showed that the sand tampan, Ornithodoros saviayni (Audouin), was capable of detecting a 5% concentration of CO₂ liberated at 1 liter/minute while still below the sand. He therefore concluded that CO₂ in the exhaled breath of the host, ca 4-5% in bovines and humans (Kellogg, 1970), was the main factor causing stimulation and attraction of the sand tampan. Of significance in Nevill's work, however, was the fact that not all of the ticks reacted to the CO₂. Also, of the ticks which were stimulated to move, their reactions were not simultaneous.

Wilson et al. (1972) also used CO₂ incorporated in a dry ice trap to investigate attraction behavior of A. americanum. Although ticks were attracted from as far away as 21.3 m, these CO₂ traps were only capable of collecting 60-80% of the ticks in the trap vicinity. Instead of going directly to the trap, many ticks were observed to take a "waiting attitude" when nearing the trap.

Alternative sources of CO₂, other than dry ice, were utilized by Sauer et al. (1974) to determine what effect CO₂ concentration had on A. americanum. Using commercially purchased cylinders of 100% CO₂ and a Y-tube olfactometer, they concluded that the intensity of the tick response increases with increasing CO₂ concentration up to ca 8%. Some ticks were once again observed to be indifferent to CO₂ stimulation regardless of concentration.

Burris (1974) examined contact and olfactory responses of the lone star tick, A. americanum, to 67 inorganic or organic chemicals soluble

in H₂O or methanol. Those chemicals containing sodium appeared to be most attractive, with statistical analysis showing certain amino acids as the chemicals eliciting the best responses. Burris also performed extensive trials utilizing a 5% concentration of CO₂ in an olfactometer system. Although results suggested an attraction response to CO₂, the averages were lower than the expected efficiency of the olfactometer. Again, questions were raised regarding factors responsible for CO₂ perception in ticks.

The use of electrophysiology as an effective means of investigating the sensory physiology of certain arthropods has been well documented. The slow and summed receptor potentials elicited by odors were first recorded by Schneider (1957) in the antenna of the male silkworm, Bombyx mori (L.). Since then, this technique has been adapted to a variety of different investigative procedures. Electroretinograms (ERG's) have been used to measure spectral sensitivity in adult bollworms, Heliothis zea (Boddie) and tobacco budworms, Heliothis virescens (F.) (Agee, 1973), Caribbean fruit flies, Anastrepha suspensa (Loew) (Agee and Park, 1975) and screwworm flies, Cochliomyia hominivorax (Coquerel) (Goodenough, 1977). Electroantennogram (EAG) responses to various compounds have been monitored in the cabbage looper, Trichoplusia ni (Hubn) (Mayer, 1973), yellowfever mosquito, Aedes aegypti (L.) (Davis and Rebert, 1972 and Davis, 1976a, 1976b, 1977), Japanese gypsy moth, Porthedria dispar japonica (Yamada et al., 1976), bark beetle, Dendroctonus frontalis Zimm, (Payne and Dickens, 1976), tobacco hornworm, Manduca sexta (Schweitzer et al., 1976), American cockroach, Periplaneta americanum (L.) (Washio and Nishino, 1976), subterranean termite, Reticulitermes lucifugus santonensis (Feytoud) (Floyd et al., 1976) and redbanded

leafroller, Argyrotaenia velutinana (Walker) (Baker and Roelofs, 1976). Even responses of the acoustic sense cells of adult bollworms and tobacco budworms have been examined electrophysiologically (Agee, 1967).

Extensive electrophysiological investigations involving ticks, however, have not been attempted. Gregson (1967) utilized electrograms to monitor the feeding behavior of D. andersoni. He observed many types of traces and associated them with different aspects of the ticks feeding pattern. Sweatman and Gregson (1970) conducted similar feeding electrograms on Hyalomma aegyptium and showed the importance of temperature on the rate of engorgement. In the area of olfactory perception, Zolotarev and Elizarov (1963) studied chemoreception in I. persulcatus. After implanting a microelectrode into the base of Haller's organ and subjecting the tick to a variety of commercially prepared repellents, they established that these chemicals effect the tick by stimulating the contact chemoreceptors in Haller's organ. Further experiments by Elizarov (1964) on Hyalomma asiaticum and I. persulcatus demonstrated that a volley of impulses of varying amplitudes could be elicited 0.5 to 1.0 seconds after introduction of various chemical irritants (odors) into Haller's organ. These volleys were usually observed to be monophasic with negative polarity but would at times be of positive polarity. More recently, Waladde and Rice (1977) obtained characteristic chemoreceptor responses to solutions of sodium chloride, adenosine triphosphate, reduced glutathion and bovine plasma from cheliceral pit sensilla of the cattle tick, B. microplus, by electrophysiological techniques. This work adds a sensory function to the chelicerae of ticks, appendages which are generally regarded as mere cutting tools.

CHAPTER III

MATERIALS AND METHODS

Tick Rearing

Experiments were conducted on adult Gulf Coast ticks, Amblyomma maculatum Koch, lone star ticks, Amblyomma americanum (L.) and American dog ticks, Dermacentor variabilis (Say). Larval and nymphal stages were reared on domestic rabbits at Oklahoma State University and stored in cardboard containers in sealed plexiglass humidity chambers containing saturated salt solutions.

Experimental Design

All ticks were preconditioned a minimum of one week prior to testing in sealed plexiglass humidity chambers. Due to the comparative nature of these studies, specific environmental and physiological conditions under which ticks were preconditioned (i.e. humidity, temperature, ambient CO₂ concentration and age) varied according to the tests performed. A photoperiod of 14 hrs daylight 10 hrs. darkness was kept constant throughout all tests.

Age Study

Ticks of 1-2, 4-6, 8-10, 12-14 and 16-18 weeks post-molt were tested after being preconditioned at room temperature ($26 \pm 0.5^{\circ}\text{C}$) to a high relative humidity ($87 \pm 4\%$) and low ambient CO₂ concentration

(422 ± 65 ppm). Humidity was kept constant using a saturated solution of K_2SO_4 (Winston and Bates, 1960). Ambient CO_2 concentration was maintained by using a 1/82 hp Fischer[®] Dyna-pump which circulated fresh air through the chamber. The level of CO_2 was checked daily with an Infrared[®] Model 755-010 spectrometer in this and subsequent studies.

Ambient CO_2 Study

Ticks of 8-10 weeks post-molt were tested after being preconditioned at $26 \pm 0.5^\circ C$ and $87 \pm 4\%$ RH to either a low (422 ± 65 ppm) or high (956 ± 78 ppm) ambient CO_2 concentration. Humidity was maintained with a saturated solution of K_2SO_4 (Winston and Bates, 1960). Low ambient CO_2 concentration was kept constant by using a Fischer[®] Dyna-pump. High ambient CO_2 levels were obtained by not circulating fresh air through the chamber, thus allowing normal room CO_2 concentrations to build-up within the chamber. Carbon dioxide levels were checked daily.

Humidity Study

Ticks of 8-10 weeks post-molt were tested after being preconditioned at $26 \pm 0.5^\circ C$ and 422 ± 65 ppm CO_2 to either a low ($65 \pm 5\%$), medium ($74 \pm 2\%$) or high ($87 \pm 4\%$) relative humidity. These respective humidity levels were obtained by using saturated solutions of $NaNO_2$, $NaCl$ and K_2SO_4 (Winston and Bates, 1960). Carbon dioxide concentration was maintained using a Fischer[®] Dyna-pump and checked daily.

Temperature Study

Ticks of 8-10 weeks post-molt were tested after being preconditioned at $87 \pm 4\%$ RH and 422 ± 65 ppm CO_2 to either a low ($21 \pm 0.8^\circ C$), medium

($26 \pm 0.5^{\circ}\text{C}$) or high ($32 \pm 0.4^{\circ}\text{C}$) temperature. The low temperature was maintained by using a standard room air conditioner and the high temperature by using a Titan[®] Model T760B portable room heater. The medium temperature was equivalent to room temperature where tests were being performed. Humidity was maintained with a saturated solution of K_2SO_4 (Winston and Bates, 1960). Carbon dioxide concentration was kept constant by using a Fischer[®] Dyna-pump and checked daily.

Experimental Set-up

A copper screen lined Faraday cage served as a holding area and provided an electrically grounded environment for tick preparation and testing. Ticks were immobilized with their ventral side up on double-sided Borden[®] mystik tape and positioned on a Pyrex[®] gas dispersion tube situated under a Bausch and Lomb[®] Model AVB-73 stereomicroscope. The dispersion tube was connected to a commercially prepared cylinder of 350 ± 18 ppm CO_2 and allowed for regulation of ambient CO_2 concentrations during testing.

An insulated (except at the tip) stainless steel microelectrode was inserted into the tick body in a ground position and served as an indifferent electrode. A similar recording microelectrode was inserted through the corium between the metatarsus and tarsus just posterior to Haller's organ (located on the dorsal tarsal surface), and connected to the input of a Grass[®] Model P15 differential AC preamplifier. The amplified (1000x) and filtered nerve impulses were then displayed on a Tektronix[®] Model 5113 dual beam storage oscilloscope. A Realistic[®] Model SA-10 solid state stereo amplifier and Realistic[®] Model Solo-5 stereo speaker were also connected to the preamplifier output and

allowed for auditory observations of tick action potentials.

Administration of CO₂ concentrations used for tick stimulation was accomplished by a fluidic gas delivery system (Pierce, 1976). The system intake was connected to a commercially prepared cylinder of 950 ± 48 ppm CO₂ and the outlet connected in circuit to the gas dispersion tube. This provided a means by which ambient CO₂ concentrations surrounding the tick could be increased from ca 350 ppm to 950 ppm at any time during testing. A 5 ft section of ¼ in Tygon® tubing was connected to an Infrared® Model 755-010 spectrometer and Linear® Model 485 strip chart recorder and positioned directly above the dispersion tube. This enabled CO₂ concentrations to be detected and recorded throughout the test period.

Data Collection and Analysis

Five female and five male ticks of each species were tested from each variable within a particular study. All ticks were purged (stimulated) ten times with a ten second burst of 950 ± 48 ppm CO₂ every two minutes. Prior to CO₂ stimulation a steady baseline was obtained, with few if any nerve impulses being observed. However, at the onset of CO₂ stimulation a rapid volley of impulses was produced which eventually returned to the initial steady baseline. This response time, in seconds, of nerve impulses produced from each burst of increased CO₂ concentration was recorded and mean values calculated.

A standard analysis of variance was run on the data and significance determined by using Duncan's Multiple Range test for comparison of treatment means.

CHAPTER IV

RESULTS

Age Study

The effects of age on mean response times to CO₂ are shown in Table 1. Average response times to CO₂ (ca 950 ppm) in A. maculatum were significantly different ($P < 0.05$) between sexes at all ages tested. Average response times in A. americanum showed significant differences ($P < 0.05$) between sexes at ages 1-2, 8-10 and 12-14 weeks post-molt. However, D. variabilis only showed significant differences ($P < 0.05$) between sexes at age 8-10 weeks post-molt.

Average response times of female ticks of each species to CO₂ (ca 950 ppm) showed A. maculatum to be significantly different ($P < 0.05$) from the other tick species throughout all ages tested. At ages 1-2 and 4-6 weeks post-molt, female response times were significantly different ($P < 0.05$) between all species. However, female response times of A. americanum and D. variabilis at ages 8-10, 12-14 and 16-18 weeks post-molt were not significantly different.

Average response times of male ticks of each species to CO₂ (ca 950 ppm) were not significant at ages 1-2, 4-6 and 8-10 weeks post-molt. At ages 12-14 and 16-18 weeks post-molt, significant differences ($P < 0.05$) in response times were observed between males of A. americanum and the other two tick species.

When one compares mean response times to CO₂ (ca 950 ppm) within

TABLE 1. Mean response time to CO₂ vs age in three species of ixodid ticks.

Age**	MEAN RESPONSE TIME (Sec.)*					
	<u>Amblyomma maculatum</u>		<u>Amblyomma americanum</u>		<u>Dermacentor variabilis</u>	
	♀	♂	♀	♂	♀	♂
1-2	43 b	31 efgh	36 cde	25 hijkl	25 hijkl	29 fghi
4-6	40 bc	28 ghij	31 efgh	26 ghijk	23 jklm	24 ijk1
8-10	53 a	25 hijkl	34 def	22 jklm	32 efg	24 ijk1
12-14	38 bcd	20 klm	25 hijkl	32 efg	29 fghi	24 ijk1
16-18	32 efg	17 m	21 klm	26 hijk	24 ijk1	19 lm

*Means followed by the same letter are not significantly different at the 5% level (Duncan).

**Weeks post-molt

a species and sex at each age, significant differences ($P < 0.05$) are observed. However, these differences are inconsistent and do not follow a particular pattern in all species.

Ambient CO₂ Study

The effects of ambient CO₂ concentration on mean response times to CO₂ are shown in Table 2. In all three species, average response times to CO₂ (ca 950 ppm) at low ambient CO₂ levels (422 ± 65 ppm) were significantly different ($P < 0.05$) between sexes. At high ambient CO₂ levels (956 ± 78 ppm), significant differences ($P < 0.05$) between sexes were observed in A. maculatum and A. americanum.

Average response times of female ticks of each species to CO₂ (ca 950 ppm) at 422 ± 65 ppm ambient CO₂ were significantly different ($P < 0.05$) between A. maculatum and the other two tick species. At 956 ± 78 ppm ambient CO₂, significant differences ($P < 0.05$) were observed between mean response times of female D. variabilis and those of A. maculatum and A. americanum.

Average response times of male ticks of each species to CO₂ (ca 950 ppm) at 422 ± 65 ppm ambient CO₂ were not significant. At 956 ± 78 ppm ambient CO₂, however, significant differences ($P < 0.05$) in male response times were observed between A. americanum and the other two tick species.

Comparison of mean response times to CO₂ (ca 950 ppm) within a species and sex, show significant differences ($P < 0.05$) occurring between 422 ± 65 ppm and 956 ± 78 ppm ambient CO₂ in all cases.

TABLE 2. Mean response time to CO₂ vs ambient CO₂ levels
in three species of ixodid ticks.

TICK SPECIES	MEAN RESPONSE TIME (Sec.)*			
	High ambient CO ₂ **		Low ambient CO ₂ ***	
	♀	♂	♀	♂
<u>Amblyomma maculatum</u>	25 c	20 de	53 a	25 c
<u>Amblyomma americanum</u>	24 c	14 f	34 b	22 cd
<u>Dermacentor variabilis</u>	18 e	18 e	32 b	24 c

*Means followed by the same letter are not significantly different at the 5% level (Duncan).

**956 ± 78 ppm

***422 ± 65 ppm

Humidity Study

The effects of humidity on mean response times to CO₂ are shown in Table 3. In all three species, average response times to CO₂ (ca 950 ppm) at the high humidity ($87 \pm 4\%$) were significantly different ($P < 0.05$) between sexes. However, only the medium ($74 \pm 2\%$) and low ($65 \pm 5\%$) humidities revealed significant differences ($P < 0.05$) between the sexes of D. variabilis and A. americanum, respectively.

Average response times of female ticks of each species to CO₂ (ca 950 ppm) at $87 \pm 4\%$ RH and $74 \pm 2\%$ RH were significantly different ($P < 0.05$) between A. maculatum and the other two tick species. At $65 \pm 5\%$ RH, significant differences ($P < 0.05$) were observed between mean response times of female D. variabilis and those of A. americanum and A. maculatum.

Average response times of male ticks of each species to CO₂ (ca 950 ppm) at $87 \pm 4\%$ RH were not significantly different. However, at $74 \pm 2\%$ RH and $65 \pm 5\%$ RH response times of male A. maculatum were significantly different ($P < 0.05$) from A. americanum and D. variabilis.

When comparing mean response times to CO₂ (ca 950 ppm) at each humidity within a species and sex, significant differences ($P < 0.05$) are observed between $87 \pm 4\%$ RH and $65 \pm 5\%$ RH in all cases. However, comparison of mean response times at $74 \pm 2\%$ RH vs $87 \pm 4\%$ or $65 \pm 5\%$ RH within a species and sex show significant differences to be inconsistent.

Temperature Study

The effects of temperature on mean response times to CO₂ are shown in Table 4. In all three species, average response times to CO₂ (ca 950 ppm) at the medium temperature ($26 \pm 0.5^\circ\text{C}$) were significantly

TABLE 3. Mean response time to CO₂ vs humidity in three species of ixodid ticks.

TICK SPECIES	MEAN RESPONSE TIME (Sec.)*					
	High humidity**		Medium humidity***		Low humidity****	
	♀	♂	♀	♂	♀	♂
<u>Amblyomma maculatum</u>	53 a	25 cd	27 c	24 cde	22 efg	20 fg
<u>Amblyomma americanum</u>	34 b	22 defg	23 def	20 fgh	21 efg	17 hi
<u>Dermacentor variabilis</u>	32 b	24 ce	22 def	19 def	15 i	15 i

*Means followed by the same letter are not significantly different at the 5% level (Duncan).

**87 ± 4%

***74 ± 2%

****65 ± 5%

TABLE 4. Mean response times to CO₂ vs temperature in three species of ixodid ticks.

TICK SPECIES	MEAN RESPONSE TIME (Sec.)*					
	High temperature**		Medium temperature***		Low temperature****	
	♀	♂	♀	♂	♀	♂
<u>Amblyomma maculatum</u>	41 b	27 d	53 a	25 de	19 h	19 h
<u>Amblyomma americanum</u>	39 b	31 c	34 c	22 fg	23 efg	20 gh
<u>Dermacentor variabilis</u>	25 de	23 ef	32 c	24 ef	28 d	28 d

*Means followed by the same letter are not significantly different at the 5% level (Duncan).

**32 ± 0.4°C

***26 ± 0.5°C

****21 ± 0.8°C

different ($P < 0.05$) between sexes. Average response times to CO_2 (ca 950 ppm) at the low temperature ($21 \pm 0.8^\circ\text{C}$) showed no significant differences occurring between sexes. At the high temperature ($32 \pm 0.4^\circ\text{C}$) significant differences ($P < 0.05$) in mean response times to CO_2 (ca 950 ppm) were observed between sexes in A. maculatum and A. americanum, but not in D. variabilis.

Average response times of female ticks of each species to CO_2 (ca 950 ppm) at $21 \pm 0.8^\circ\text{C}$ were significantly different ($P < 0.05$) in all cases. Average response times of female ticks of each species at $32 \pm 0.4^\circ\text{C}$ showed significant differences ($P < 0.05$) between D. variabilis and the other two tick species. At $26 \pm 0.5^\circ\text{C}$, significant differences ($P < 0.05$) in response times were observed between females of A. maculatum and those of A. americanum and D. variabilis.

Average response times of male ticks of each species to CO_2 (ca 950 ppm) at $32 \pm 0.4^\circ\text{C}$ were significantly different ($P < 0.05$) in all cases. At $26 \pm 0.5^\circ\text{C}$, mean response times of male A. maculatum were significantly different ($P < 0.05$) from male A. americanum. Average response times to CO_2 (ca 950 ppm) in males at $21 \pm 0.8^\circ\text{C}$ showed significant differences ($P < 0.05$) between D. variabilis and the other two tick species.

When comparing mean response times to CO_2 (ca 950 ppm) at each temperature within a species and sex, significant differences ($P < 0.05$) are observed between $32 \pm 0.4^\circ\text{C}$ and $21 \pm 0.8^\circ\text{C}$ in A. maculatum and A. americanum, respectively. Comparison of mean response times at $26 \pm 0.5^\circ\text{C}$ in A. maculatum and A. americanum showed significant differences to be inconsistent. Significant differences in mean response times were also inconsistent when comparing D. variabilis at each temperature.

CHAPTER V

DISCUSSION AND CONCLUSIONS

Results of this comparative study were interesting in that a wide and varied response was observed from each tick species and between sexes within a species, to the controlled variables. One of the most striking differences in tick response time to CO₂ was observed in 8-10 week post-molt A. maculatum females. The mean response time was 53 seconds, approximately 20 seconds longer than females of the other two species. This increased stimulation to CO₂ may be one factor involved in the selection of a feeding site on cattle, its primary host as an adult tick. This particular tick species is normally found on the ears of cattle, where the concentration of CO₂ is very substantial due to normal respiration. In contrast, A. americanum and D. variabilis do not seem to prefer any single feeding location as evidenced by observations of their natural hosts under field conditions. Differences in mean response times between sexes within a particular species were usually consistent, with female response times being greater than males. This finding agrees with field tests conducted by Wilson (1972) in which greater numbers of female A. americanum ticks were collected from CO₂ baited traps than males.

Another interesting aspect of this study was the effect on tick response time when they were preconditioned to either low or high ambient concentrations of CO₂ prior to testing. When ticks were

conditioned to low ambient CO₂ concentration (ca 400 ppm), the response time was always greater for all tick species and sexes when compared to high preconditioned CO₂ concentration (ca 950 ppm). This suggests that the background CO₂ concentration may be an important factor in the tick's ability to sense and locate a source of CO₂ under field conditions. In the field, normal diurnal CO₂ concentration is approximately 350-450 ppm with nocturnal concentrations reaching 850-950 ppm (Gearhart, unpublished data). Field work by Wilson (1972) on A. americanum showed a 55% retrap of adult ticks during daylight hours, but only a 5% retrap at night. This decrease in trapping efficiency could possibly be explained by our findings as well as other factors such as decreased nocturnal temperatures.

Another parameter measured was the effect of different humidities on tick response time to CO₂. This study was conducted under conditions equivalent to diurnal ambient CO₂ concentrations with a high concentration of CO₂ (ca 950 ppm), simulating a tick trap, being used to stimulate the tick. In all cases, as humidity increased, response time increased within a particular sex and species. In nature, tick survival and longevity is enhanced when the microenvironment humidity is constantly greater than 85% (Hair et al., 1975). Observations of tick (A. americanum) collection in the field, over a period of several years, indicate that the most ideal tick trapping areas are in shaded, leaf littered, forest floor habitats or high grass and brushy ecotone habitats (Wilson, 1972). The microenvironment humidity is normally high in these areas and thus, an ideal habitat for tick survival.

Tick behavior and seasonal activity is also affected by temperature (Wilson, 1972). The purpose of our temperature study was not to

elucidate the high and low temperature activity thresholds, and their effect on tick response to CO₂, but rather, to determine what effect temperature that are most prevalent during the tick activity season might have on tick response time to CO₂. In general, the differences in response times to CO₂ at the different temperatures were affected by the tick species and the sex of ticks of a species. Amblyomma americanum was most responsive to CO₂ at the highest temperature (ca 32°C) whereas, the other two tick species were most responsive to CO₂ at the medium temperature (ca 26°C). Wilson, in his work with A. americanum, found that CO₂ trapping efficiency increased with increasing temperature up to a peak of ca 28°C. These observed differences may, therefore, indicate approximate temperatures at which CO₂ traps would be most effective for tick trapping of a particular species.

Our findings indicate that species, sex, age, ambient CO₂ concentration, humidity and temperature do play a role in tick perception of CO₂. These results may, therefore, lend themselves to future investigations involving trap-retrap studies or tick population density surveys utilizing CO₂ traps. Results may also be of importance in rearing and preconditioning of laboratory ticks prior to use in field studies. As with all electrophysiological investigations of this type, field tests of an identical nature need to be performed before a true understanding of CO₂ perception can be obtained.

SELECTED BIBLIOGRAPHY

- Agee, H. R. 1967. Response of acoustic sense cell of the bollworm and tobacco budworm to ultrasound. J. Econ. Ent. 60(2): 366-369.
- Agee, H. R. 1973. Spectral sensitivity of the compound eyes of field-collected adult bollworms and tobacco budworms. Ann. Ent. Soc. Amer. 66(3): 613-615.
- Agee, H. R. and M. L. Park. 1975. Use of the electroretinogram to measure the quality of vision of the fruit fly. Environmental Letters. 10(2): 171-176.
- Baker, T. C. and W. L. Roelofs. 1976. Electroantennogram response of the male moth, Argyrotaenia velutinana to mixtures of sex pheromone components of the female. J. Insect Physiol. 22: 1357-1364.
- Balashov, Y. S. 1972. Bloodsucking ticks (Ixodoidae) - Vectors of diseases of man and animals. Misc. Pub. of the Ent. Soc. Amer. 8(5): 193-213.
- Bruce, W. A. 1971. Posterior capsule of Haller's organ in the lone star tick, Amblyomma americanum (Acari: Ixodidae). Fla. Entomol. 54(1): 65-72.
- Burris, E. 1974. Laboratory tests of candidate chemical attractants for ticks. M.S. Thesis. Oklahoma State University.
- Chow, T. S. and C. H. Wang. 1975. External structure of Haller's organ of the Ixodes tick. Bull. Inst. Zool., Academia Sinica. 14(1): 9-17.
- Chu-Wang, I. and R. C. Axtell. 1973. Comparative fine structure of the claw sensilla of a soft tick, Argas (Persicargas) arboreus Kaiser, Hoogstraal and Kohls, and a hard tick, Amblyomma americanum (L.). J. Parasitol. 59(3): 545-555.
- Davis, E. E. and C. S. Rebert. 1972. Elements of olfactory receptor coding in the yellowfever mosquito. J. Econ. Ent. 65(4): 1058-1061.
- Davis, E. E. 1976a. A receptor sensitive to oviposition site attractants on the antennae of the mosquito, Aedes aegypti. J. Insect Physiol. 22: 1371-1376.

- Davis, E. E. 1976b. Lactic acid-sensitive receptors on the antennae of the mosquito, Aedes aegypti. J. comp. Physiol. 105: 43-54.
- Davis, E. E. 1977. Response of the antennal receptors of the male Aedes aegypti mosquito. J. Insect Physiol. 23:613-617.
- Dethier, V. G. 1957. The sensory physiology of bloodsucking arthropods. Exp. Parasitol. 6: 68-122.
- Elizarov, Y. A. 1964. Investigation of chemoreception of insects and ticks. I. Electrical activity of chemoreceptors of Haller's organ during action of repellents. Nauchnyy Doklady Vvsshey Shkoly, Biologicheskoye Nauki. 2: 55-59.
- Erickson, R. P. 1963. Sensory neural patterns and gustation. In: Olfaction and Taste, ed. by Y. Zotterman, Pergamon Press, Oxford. pp. 205-213.
- Floyd, M. A., D. A. Evans and P. E. Howse. 1976. Electrophysiological and behavioural studies on naturally occurring repellents to Reticulitermes lucifugus. J. Insect Physiol. 22: 697-701.
- Foelix, R. F. and R. C. Axtell. 1972. Ultrastructure of Haller's organ in the tick, Amblyomma americanum (L.). Z. Zellforsch. 124: 275-292.
- Garcia, R. 1962. Carbon dioxide as an attractant for certain ticks (Acarina: Argasidae and Ixodidae). Ann. Ent. Soc. Amer. 55: 605-606.
- Garcia, R. 1965. Collection of Dermacentor andersoni (Stiles) with carbon dioxide and its application in studies of Colorado tick virus. Amer. J. Trop. Med. Hyg. 14: 1090-1093.
- Garcia, R. 1969. Reaction of the winter tick, Dermacentor albipictus (Packard), to CO₂. J. Med. Ent. 6: 206.
- Goodenough, J. L., D. D. Wilson and H. R. Agee. 1977. Electroretinographic measurements for comparison of visual sensitivity of wild and mass-reared screwworm flies, Cochliomyia hominivorax (Diptera: Calliphoridae). J. Med. Ent. 19(3): 309-312.
- Gregson, J. D. 1967. Electrical observations of tick feeding in relation to disease transmission. Proceedings of 2nd Int. Congress of Acarology. 329-339.
- Hair, J. A., J. R. Sauer and K. A. Durham. 1975. Water balance and humidity preference in three species of ticks. J. Med. Ent. 12(1): 37-47.
- Hindle, E. and G. Merriman. 1912. The sensory perceptions of Argas persicus (Oken). Parasit. 5: 203-216.

- Homsher, P. J. and D. W. Sonenshine. 1975. Scanning electron microscopy of ticks for systematic studies: Fine structure of Haller's organ in ten species of Ixodes. *Trans. Amer. Micros. Soc.* 94(3): 368-374.
- Huber, F. 1967. Central control of movements and behavior of invertebrates. In: *Invertebrate Nervous Systems*, ed. by C. A. G. Wiersma, University of Chicago, Chicago, Ill. pp. 333-352.
- Kaissling, K. E. 1971. Insect olfaction. In: *Handbook of Sensory Physiology. IV. Olfaction*, ed. by L. M. Beidler, Springer, New York. pp. 351-431.
- Kellogg, F. E. 1970. Water vapor and carbon dioxide receptors in *Aedes aegypti*. *J. Insect Physiol.* 16: 99-108.
- LaHille, F. 1905. Contributions a l'etude des Ixodes de la Republique Argentine. *Anales del Minist. de Agric. Seccion de Zootecnia.* II: 107-109.
- Lees, A. D. 1948. The sensory physiology of the sheep tick, *Ixodes ricinus* (L.). *J. Exp. Biol.* 25(2): 149-207.
- Mayer, M. S. 1973. Electrophysiological correlates of attraction in *Trichoplusia ni*. *J. Insect Physiol.* 19: 1191-1198.
- Miles, V. I. 1968. A carbon dioxide bait trap for collecting ticks and fleas from animal burrows. *J. Med. Ent.* 5: 491-495.
- Nevill, E. M. 1964. The role of carbon dioxide as a stimulant and attractant to the sand tampan, *Ornithodoros savignyi* (Audouin). *Onderstepoort J. Vet. Res.* 31: 59-68.
- Nuttall, G. H. F., W. F. Cooper and L. E. Robinson. 1908. On the structure of Haller's organ in the Ixodoidea. *Parasit.* 1:238-243.
- Payne, T. L. and J. C. Dickens. 1976. Adaption to determine receptor system specificity in insect olfactory communications. *J. Insect Physiol.* 22: 1569-1572.
- Pierce, S. K. 1976. A gas chromatographic determination of metabolic profiles in human breath and urine. M.S. Thesis. Oklahoma State University.
- Rechav, Y., S. Terry, M. M. Knight and R. H. M. Cross. 1977. Chemoreceptor organs used in detection of pheromone(s) of the tick *Amblyomma hebraeum* (Acarina: Ixodidae). *J. Med. Ent.* 14(4): 395-400.
- Roeder, K. D. 1970. Episodes in insect brains. *American Scientist.* 58: 378-389.

- Sactor, B., M. Hutchinson and P. Granett. 1948. Biology of the lone star tick in the laboratory. *J. Econ. Ent.* 41(2): 296-301.
- Sauer, J. R. and J. A. Hair. 1971. Water balance in the lone star tick (Acarina: Ixodidae): The effects of relative humidity and temperature on weight changes and total water content. *J. Med. Ent.* 8(5): 479-485.
- Sauer, J. R., J. A. Hair and M. S. Houts. 1974. Chemoattraction in the lone star tick (Acarina: Ixodidae). 2. Responses to various concentrations of CO₂. Journal article #2683 Agric. Exp. Stn. Oklahoma State University.
- Schneider, D. 1957. Electrophysiological investigations of chemo- and mechanoreceptors of the antenna of the silk moth, Bombyx mori L. *Ztschr. f. Vergl. Physiol.* 40: 8-41.
- Schweitzer, E. S., J. R. Saues and J. G. Hildebrand. 1976. Ontogeny of electroantennogram response in the moth, Manduca sexta. *J. Insect Physiol.* 22: 955-960.
- Slifer, E. H. 1970. The structure of arthropod chemoreceptors. *Ann. Rev. Entomol.* 15: 121-142.
- Sweatman, G. K. and J. D. Gregson. 1970. Feeding electrograms of Hyalomma aegyptium ticks at different temperatures. *J. Med. Ent.* 7(5): 575-584.
- Waladde, S. M. 1977. The sensory nervous system of the adult cattle tick Boophilus microplus (Canestrini) Ixodidae. Part II. Scanning electron microscopy. *J. Aust. ent. Soc.* 16: 73-79.
- Waladde, S. M. and M. J. Rice. 1977. The sensory nervous system of the adult cattle tick Boophilus microplus (Canestrini) Ixodidae. Part III. Ultrastructure and electrophysiology of cheliceral receptors. *J. Aust. ent. Soc.* 16: 441-453.
- Washio, H. and C. Nishino. 1976. Electroantennogram responses to the sex pheromone and other odours in the American cockroach. *J. Insect Physiol.* 22: 735-741.
- Wilkinson, P. R. 1953. Observations on the sensory physiology and behavior of larvae of the cattle tick, Boophilus microplus (Can.). (Ixodidae). *Aust. J. Zool.* 1(3): 345-356.
- Wilson, J. G. 1972. Responses of the lone star tick, Amblyomma americanum (Linnaeus), to carbon dioxide stimulation. M.S. Thesis. Oklahoma State University.
- Wilson, J. G., D. R. Kinzer, J. R. Sauer and J. A. Hair. 1972. Chemoattraction in the lone star tick (Acarina: Ixodidae): I. Response of different developmental stages to carbon dioxide administered via traps. *J. Med. Ent.* 9(3): 245-252.

Winston, P. W. and D. H. Bates. 1960. Saturated solutions for the control of humidity in biological research. *Ecology*. 41(1): 232-237.

Yamada, M., T. Saito, K. Katagiri, S. Iwaki and S. Marumo. 1976. Electroantennogram and behavioural responses of the gypsy moth to enantiomers of disparture and its trans analogues. *J. Insect Physiol.* 22: 755-761.

Zolotarev, E. K. and Y. A. Elizarov. 1963. Studies of chemoreception in insects and ticks. Location of the chemoreceptors in the tick, Ixodes persulcatus P. Sch. *Vestnik Moskovskogo Universitetus. Seriya VI, Biologiya Pochevedinie.* 18(1): 7-9.

VITA 2

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